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The scaling and temperature dependence of vertebrate metabolism

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Body size and temperature are primary determinants of metabolic rate, and the standard metabolic rate (SMR) of animals ranging in size from unicells to mammals has been thought to be proportional to body mass (M) raised to the power of three-quarters for over 40 years. However, recent evidence from rigorously selected datasets suggests that this is not the case for birds and mammals. To determine whether the influence of body mass on the metabolic rate of vertebrates is indeed universal, we compiled SMR measurements for 938 species spanning six orders of magnitude variation in mass. When normalized to a common temperature of 38 °C, the SMR scaling exponents of fish, amphibians, reptiles, birds and mammals are significantly heterogeneous. This suggests both that there is no universal metabolic allometry and that models that attempt to explain only quarterpower scaling of metabolic rate are unlikely to succeed.

Keywords: scaling; allometry; metabolic rate; body mass; body temperature

1. INTRODUCTION

The study of the allometric relationship between metabolic rate and body mass is currently in the midst of a renaissance (see Agutter & Wheatley 2004 for a recent review). Much of the impetus for this renewed interest can be attributed to the publication of models (Banavar et al. 1999, 2002; West et al. 1997, 1999) seeking to explain the widely accepted observation that metabolic rate is proportional to body mass raised to the three-quarter power $(M^{0.75})$ in organisms ranging in size from unicells to large mammals (Hemmingsen 1960). However, it is now clear that the basal metabolic rate of endotherms scales with an exponent less than three-quarters (White & Seymour 2003; McKechnie & Wolf 2004), and the empirical and mathematical support for a universal metabolic allometry have been questioned (Dodds et al. 2001; Bokma 2004; Kozlowski & Konarzewski 2004). Although many physiological rates and time-scale with exponents that are close to simple multiples of one-quarter (Savage et al. 2004), many others do not, and recognition of this has led to

The electronic supplementary material is available at http://dx.doi. org/10.1098/rsbl.2005.0378 or via http://www.journal.royalsoc.ac. uk. attempts to link cellular and whole-animal metabolism in a phenomenological description of scaling exponent heterogeneity (Darveau *et al.* 2002; Hochachka *et al.* 2003). However, before the nonisometric scaling of metabolic rate can be adequately explained, it must first be characterized.

2. MATERIAL AND METHODS

We compiled values for body mass (M) and standard metabolic rate (SMR) from the peer-reviewed literature and located measurements for species ranging in mass from 154 mg to 138 kg (see Electronic Appendix). Data were separated into five groups for analysis: fish, amphibians, reptiles, birds and mammals. All available data were used for determination of the temperature dependence of metabolism, and species' averages of temperaturenormalized data were used for determination of scaling exponents. This was necessary to avoid over-representation of frequently studied species in the scaling relationships. The temperature normalization was undertaken according to both van't Hoff (Q_{10}) and Boltzmann-Arrhenius principles (UTD normalization: Gillooly et al. 2001). The van't Hoff principle has long been the mainstay for the description of the temperature dependence of metabolism and, at least for mammals (which have the largest dataset), accounts for more of the temperature-associated variation in metabolism than does UTD normalization (White & Seymour 2003). However, for the poikilothermic species in the present dataset, ln(SMR) is not linearly related to temperature; temperature is not a significant factor in a model that described ln(SMR) in terms of the following fixed factors: group (fish, reptile, amphibian) + temperature + 1000/temperature + $\ln(M)$ + group × temperature + group $\times 1000$ /temperature + group ln(\dot{M}) and a random factor: species; all other factors and interactions were significant (p < 0.05). Thus, while there is no theoretical justification for preferring either van't Hoff or UTD normalization (Clarke 2004), UTD normalization provides a better description of the temperature dependence of metabolism for the poikilothermic species, and both van't Hoff and UTD normalization were undertaken.

The data were not 'binned' (Savage et al. 2004) to account for a perceived over-representation of small species; the distribution of vertebrate body sizes is considered to represent an important component of diversity, rather than a bias that must be accounted for. Furthermore, the body mass composition of the most wellrepresented group in the dataset (mammals) closely matches that of extant species (Blackburn & Gaston 1998): when divided into size classes spanning 0.5 log units of body mass, species weighing less than 32 g are under-represented by 0.7-6.3% (mean: 2.6%), species in the range 32 g to 10 kg are over-represented by 1.7-6.8% (mean: 3.5%), and species >10 kg are under-represented by 0.2-2.8% (mean: 1.3%). The distribution of body masses of the sample of birds is similarly close to that of extant species (Blackburn & Gaston 1994), with differences of -2.8% to 6.2%, but comparable body mass distribution data are not available for fish, reptiles and amphibians. Pough's (1980) body mass data, estimated from snout-vent length measurements of 5339 species, suggest that body mass classes within the present amphibian dataset are appropriately distributed (differences of -3.3 to 2.7%), while large (>100 g) reptiles are over-represented by 14% and small ones (<100 g) are under-represented by 1.6–5.7%.

SMRs for endotherms were included only if measured under basal conditions (McNab 1997; Frappell & Butler 2004) and if body temperature was recorded. Data for lineages for which basal conditions are unlikely to be achieved (Artiodactyla, Lagomorpha, Macropodidae, Soricidae) were excluded for reasons discussed elsewhere (White & Seymour 2003, 2005). SMRs for poikilotherms were included only if measured under standard resting conditions at a known ambient temperature (T_a , range: 1–45 °C). However, the MR elevation that follows feeding for endotherms (White & Seymour 2005) is smaller and shorter in duration than that of reptiles (Secor & Diamond 1997), whose MR continues to decline following feeding: long-term fasted animals have MRs significantly lower than postabsorptive ones (Bedford & Christian 2001). Care was therefore taken to ensure, as far as possible, that data for poikilotherms were included only if animals were fasted for sufficiently long to ensure a postabsorptive state, but not so long that MR had declined significantly below postabsorptive levels.

Within each group, normalization to a common T_a (38 °C for endotherms; 38 °C and 20 °C for poikilotherms) was first accomplished by iteratively simultaneously solving for Q_{10} and the scaling exponent (White & Seymour 2005). The Q_{10} used for temperature normalization was calculated as the ordinary least squares (OLS) Table 1. Parameter estimates for the scaling relationship between standard metabolic rate (SMR, mL O₂ h⁻¹) and body mass (*M*, g) normalized to 38 °C; SMR= aM^b . (*n* is the number of species (number of measurements used for Q₁₀ determination in parentheses). Q₁₀ values in parentheses are 95% confidence limits. Allometric coefficient (*a*) values are for 38 °C (and 20 °C in parentheses for poikilotherms). 95% CI values for the allometric exponents (*b*) are in parentheses. OLS is the ordinary least squares regression, RMA is the reduced major axis regression calculated by passing the RMA exponent ($=b_{OLS}/\sqrt{[r^2]}$) through the bivariate log–log mean.)

			OLS		RMA		
	n	Q_{10}	a	Ь	a	Ь	r^2
fish	82 (1107)	1.65 (1.56–1.74)	0.22 (0.09)	0.88 (0.06)	0.19 (0.08)	0.91 (0.06)	0.92
amphibians	158 (682)	2.21 (2.05-2.37)	0.34 (0.08)	0.88 (0.05)	0.30 (0.07)	0.94 (0.05)	0.88
reptiles	159 (483)	2.44 (2.25-2.64)	0.55 (0.11)	0.76 (0.04)	0.46 (0.09)	0.80 (0.04)	0.91
birds	83 (83)	2.09 (1.45-3.05)	6.07	0.64 (0.03)	5.36	0.66 (0.03)	0.95
mammals	456 (469)	2.81 (2.44-3.24)	4.98	0.676 (0.013)	5.10	0.69 (0.013)	0.96



slope of the relationship between $\ln(mass-independent MR)$ and T_a ;

mass-independent MR was calculated using a scaling exponent

3. RESULTS AND DISCUSSION

Our analysis of 2824 measurements of 938 species of vertebrate expands upon the 424 measurements presented by Gillooly et al. (2001) and reveals significant allometric exponent heterogeneity (figure 1). With the exception of fish, Q_{10} falls between 2 and 3 for all groups (table 1). The Q_{10} value for fish is significantly lower than that for all other groups except birds, despite being relatively uncertain. Mammals have the highest Q_{10} (2.8), which is significantly higher than fish and amphibians (table 1). The SMR scaling exponent for reptiles is not influenced by the overrepresentation of species weighing more than 100 g: the scaling exponent for small (<100 g) reptiles is not significantly different from that of large (>100 g) ones (ANOVA mass \times size class interaction, p = 0.67 for van't Hoff normalized SMR, p=0.65 for Boltzmann-Arrhenius normalized SMR). When normalized to a common temperature of 38 °C using van't Hoff (Q_{10}) principles, fish and amphibians do not have

although RMA slopes tended to be higher than OLS ones (table 1).

Figure 1. Scaling relationships between SMR normalized to a body temperature of 38 °C and body mass for mammals, birds, reptiles, amphibians and fish. Q_{10} values used for temperature normalization are presented in table 1, as are OLS (pictured) and RMA regression parameters.

Table 2. Scaling relationship between standard metabolic rate (SMR, mL O₂ h⁻¹) and body mass (*M*, g) normalized to 38 °C (20 °C in parentheses) according to van't Hoff (Q_{10}) and Boltzmann–Arrhenius (UTD: Gillooly *et al.* 2001) principles; SMR= aM^b .

group	Q_{10}	UTD
fish	$0.224 M^{0.879}$ (0.091 $M^{0.879}$)	$\begin{array}{c} 0.213 \ M^{0.879} \\ (0.092 \ M^{0.879}) \end{array}$
amphibians	$\begin{array}{c} 0.335 \ M^{0.884} \\ (0.081 \ M^{0.884}) \end{array}$	$\begin{array}{c} 0.302 \ M^{0.884} \\ (0.081 \ M^{0.884}) \end{array}$
reptiles	$\begin{array}{c} 0.560 \ M^{0.768} \\ (0.110 \ M^{0.768}) \end{array}$	$\begin{array}{c} 0.536 \ M^{0.769} \\ (0.111 \ M^{0.769}) \end{array}$
birds mammals	$5.773 \ M^{0.644} \\ 4.648 \ M^{0.678}$	$5.801 \ M^{0.644} \\ 4.668 \ M^{0.678}$

significantly different SMRs and scale with exponents significantly higher than reptiles (table 1). The exponent for reptiles is significantly higher than that for birds and mammals. The normalization of poikilotherm data to 20 °C did not change any exponents, but influenced only the allometric coefficients (*a*, where SMR= aM^b : table 1). Temperature normalization according to van't Hoff and Boltzmann– Arrhenius principles produced scaling exponents that differed by only $\pm 0.03\%$ (table 2). The largest biology letters difference between the two temperature normalization procedures was in the scaling coefficient of the poikilothermic groups at 38 °C, which were 2.2%, 4.5% and 1.9% lower for fish, amphibians and reptiles, respectively, when normalization was conducted according to Boltzmann-Arrhenius rather than van't Hoff principles (table 2). The finding of significant allometric exponent heterogeneity fails to support Hemmingsen's (1960) and Gillooly et al.'s (2001) suggestion that the relationship between metabolic rate and body mass is uniform across all organisms. The exponent heterogeneity observed between groups, and that observed between metabolic levels within groups (White & Seymour 2005), suggest that the search for an explanation for the non-isometric scaling of metabolic rate is unlikely to end with models that explain only quarter-power scaling.

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